



### Staining Protocol for Immunocytochemistry

**Note:** The SenTraGor™ Protocol for Immunocytochemistry is a hybrid histochemical/immunohistochemical assay. To produce a consistent and specific signal it is required to use a primary anti-biotin antibody and a secondary antibody against your primary anti-biotin antibody and follow suggested steps.

For troubleshooting regarding your staining please check our SenTraGor™ Troubleshooting Guide on our website, [www.sentrageotech.com](http://www.sentrageotech.com), under Troubleshooting page.

## 1. Preparation of the biological material

### Materials:

- 1.1 Cells (from aspiration or cell culture)
- 1.2 Coverslips and cover glass
- 1.3 Glass beaker
- 1.4 10x Phosphate Buffered Saline (PBS) stock solution: 1.37 M NaCl, 27 mM KCl, 100 mM Na<sub>2</sub>HPO<sub>4</sub>, 18 mM KH<sub>2</sub>PO<sub>4</sub>, pH 7.4
- 1.5 Preparation of fixative media 1-5% (w/v) Paraformaldehyde/PBS: Dissolve 1-5 gr of paraformaldehyde (PFH) in 100 ml of PBS in a glass beaker. Heat and stir the mixture until it becomes transparent. Let the solution cool down and adjust pH to 7.4 (**Notes 4.1-4.3**)
- 1.6 Incubation chambers for coverslips
- 1.7 Positively charged glass slides
- 1.8 Thin edged forceps.

### Procedure:

Mount cells on coverslips and fix them in 1-5% (w/v) paraformaldehyde/PBS solution for 5 min at RT. Then wash three times (approx. 1 min) with PBS (**Notes 4.1**).

## 2. Preparation of SenTraGor™ reagent solution

### Materials:

- Vial with SenTraGor™ reagent
- 100% EtOH
- Parafilm

### Procedure:

- 2.1 Add 3.5-3.75 ml (20 mg SenTraGor™) or 7-7.5 ml (40 mg SenTraGor™) or 14-15 ml (80 mg SenTraGor™) 100% EtOH in the vial with the reagent and cover it with its cap and parafilm (**Notes 4.1, 4.2 and 4.4**)
- 2.2 Incubate at 56°C in a waterbath for 120 min until the reagent is completely dissolved. Store at RT (**Note 4.5**).

## 3. SenTraGor™ staining method

### Materials:

- Syringe
- 13 mm filter, membrane 0.22 µm
- Soft paper (dry or dipped in ethanol)
- Anti-biotin antibody
- Secondary antibody, HRP conjugated, against your anti-biotin antibody
- Detection system HRP DAB kit



- Hematoxylin
- 10x Tris Buffered Saline (TBS) stock solution: 1.5 M NaCl, 0.1 M Tris-HCl, pH 7.4. Store at 4°C (**Note 4.6**)
- 0.5% Triton X/TBS: 0.5 ml Triton X diluted in 99.5 ml TBS
- Mounting media (ready to use or 40% Glycerol in TBS)
- Light Microscope
- Fluorescent microscope (**optional, Note 4.7**).

### Procedure:

- 3.1 Wash coverslips x1 in TBS for 5 min at RT
- 3.2 Block endogenous hydrogen peroxidase, according to the instructions included in the Detection system HRP DAB kit, in dark conditions (**Notes 4.8, 4.18**)
- 3.3 Wash x2 in TBS for 30 sec and x1 for 5 min at RT
- 3.4 Wash x1 in 50% EtOH for 5 min at RT
- 3.5 Wash x1 in 70% EtOH for 5 min at RT (**Notes 4.9**)
- 3.6 Incubate with SenTraGor™ reagent at RT. Place coverslip with cells (using thin edged forceps) on a clean glass slide (face up). A drop of prepared reagent is placed on coverslip with cells, through a syringe attached with a 13 mm filter and membrane 0.22 µm. Then a cover glass is placed on the coverslip (using thin edged forceps) (**Notes 4.8, 4.10**) (see Figures 1 & 2)

**Figure 1**

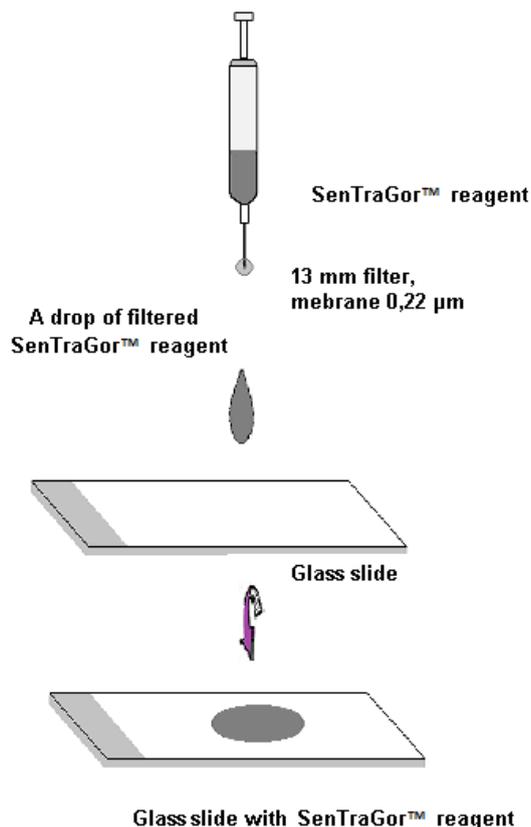
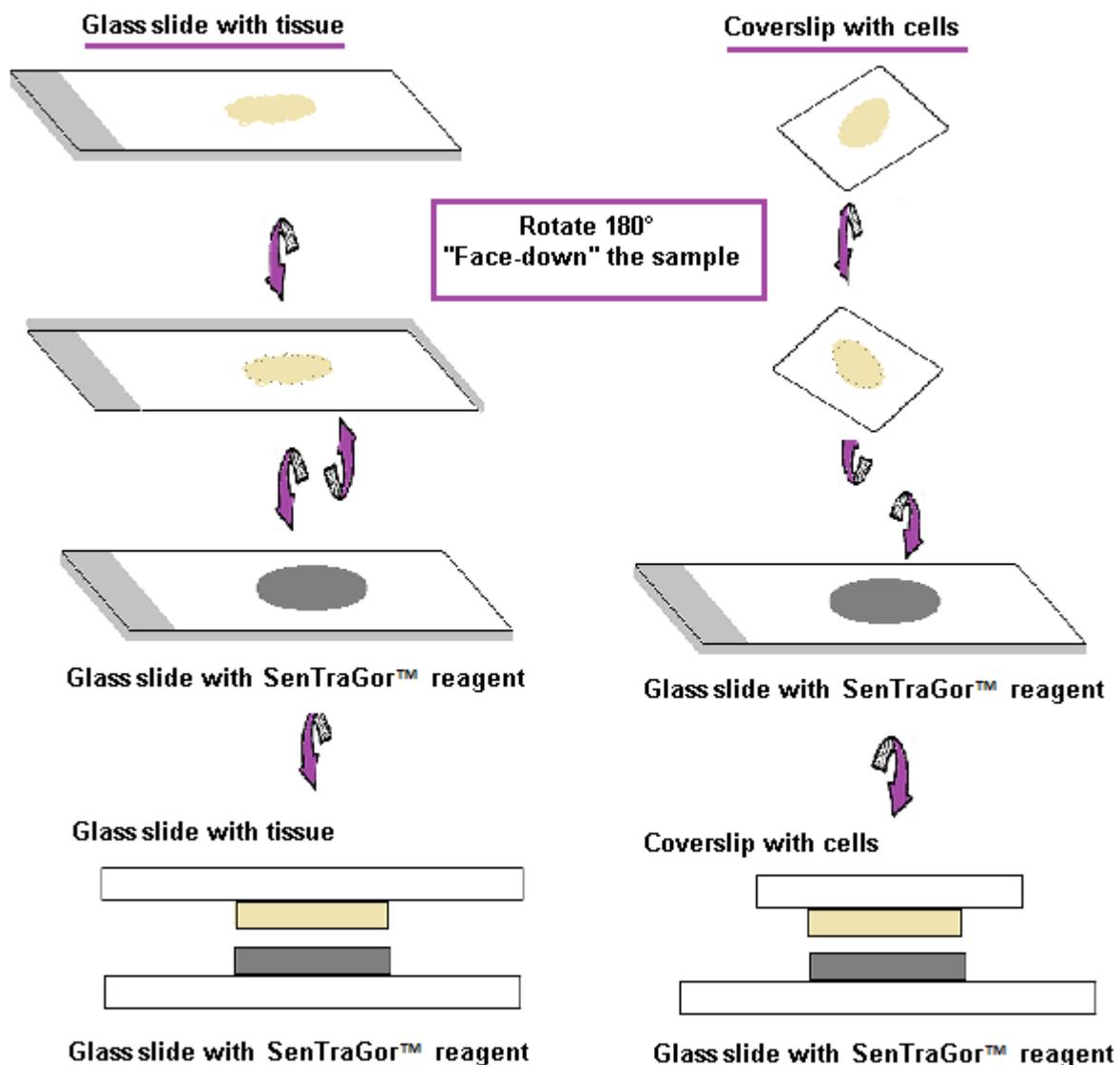




Figure 2

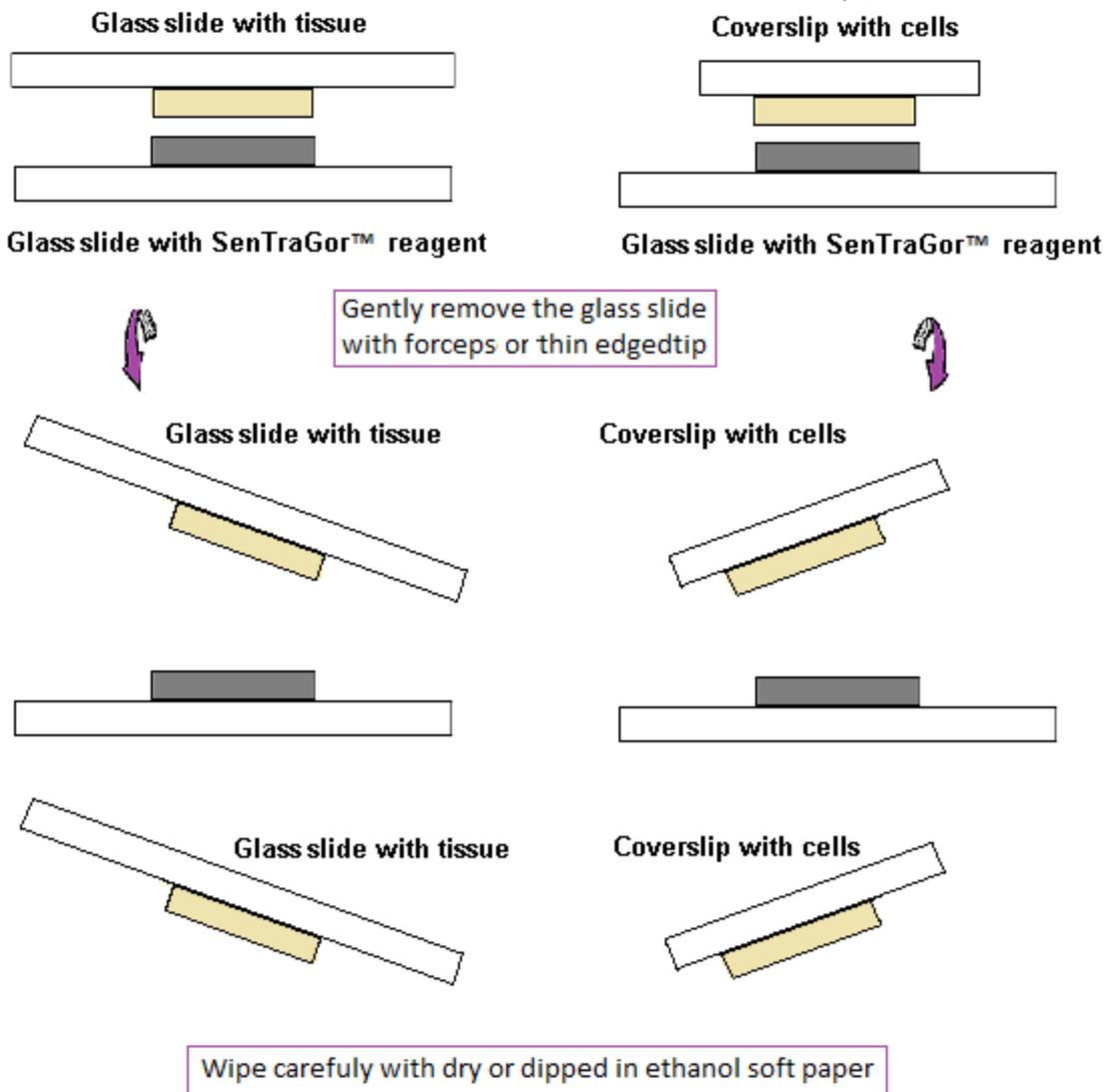


3.7 Incubate 3-8 min at room temperature or 8-10 min at 37°C (average time). The incubation time is related to the type of the sample. It is recommended to begin with 8 min incubation at RT. Continuous monitoring of the staining reaction (starting from the 3rd min) under the light microscope until detection of the signal is necessary in order to identify the optimum incubation time for each sample type (Notes 4.11-4.13)



- 3.8 Remove gently the cover glass using thin edged forceps and clean excess SenTraGor™ reagent with soft paper (ideally dipped in ethanol) (see Figure 3) (**Note 4.14**)

Figure 3



- 3.9 Wash x2 in 50% EtOH for 5 min at RT (**Note 4.14**)  
3.10 Repeat washing x2 in fresh 50% EtOH for 5 min at RT (**Note 4.14**)  
3.11 Wash x2 in TBS for 30 sec and x1 for 5 min at RT  
3.12 Incubate with 0.5% Triton X/TBS for 3 min at RT  
3.13 Wash x1 in TBS for 5 min at RT



# SenTraGor™

Antibody - enhanced detection of Senescent cells

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- 3.14 Incubate with the primary anti-biotin antibody, diluted in TBS according with your standard procedure (indicative dilution: 1/300-1/500), for 60 min at 37°C or overnight at 4°C (**Note 4.15 and 4.16**)
- 3.15 Wash x3 in TBS for 5 min at RT
- 3.16 Incubate with the secondary antibody against your anti-biotin antibody for 1h at RT
- 3.17 Wash x3 in TBS for 5 min at RT
- 3.18 Proceed according to the instructions included in your Detection system HRP DAB kit
- 3.19 Wash x2 in TBS for 30 sec and x1 for 5 min at RT
- 3.20 Apply DAB Chromogen according to the instructions included in the Detection system HRP DAB kit. The staining reaction is monitored under the light microscope until detection of the dark brown signal at RT
- 3.21 Wash in tap water for 5 min at RT
- 3.22 Counterstain with Hematoxylin (**Note 4.17**)
- 3.23 Wash in tap water for 5 min at RT
- 3.24 Apply permanent mounting media
- 3.25 Observe under the light microscope (**Notes 4.12 and 4.13**).

## 4. Technical Notes

- 4.1 Follow accurately all safety regulations (wear gloves, mask and glasses) during manipulations and waste disposal instructions when disposing waste materials.
- 4.2 Prepare all solutions using deionized water (unless otherwise indicated).
- 4.3 Preparation of the Paraformaldehyde/PBS solution must be performed in a fume hood to avoid any contact with fumes. Preferably always prepare a fresh solution before the experiments.
- 4.4 The ideal concentration depends on the examined biological material and its processing and can be determined as follows: start with 3.5 ml (20 mg SenTraGor™) or 7 ml (40 mg SenTraGor™) or 14 ml (80 mg SenTraGor™) volume of 100% Ethanol. If non-specific (“dirt background”) reaction of the reagent is observed adjust final volume to 3.75 ml (20 mg SenTraGor™) or 7.5 ml (40 mg SenTraGor™) or 15 ml (80 mg SenTraGor™), respectively.
- 4.5 Store the SenTraGor™ reagent in a non-light absorbing and airtight container at room temperature for up to 2 months. Upon longer intervals between experiments preferentially prepare a fresh solution of the dye. During the entire process the dye container must be air tightly sealed to prevent evaporation of ethanol, which in turn leads to precipitation of the saturated dye solution in cells.
- 4.6 Instead of TBS you can use PBS all the way.
- 4.7 The Fluorescent Microscope can be used in control experiments. Lipofuscin that accumulates in senescent cells is well known to exhibit autofluorescent properties that are quenched by the current SenTraGor™ reagent staining. Mount the sample in 40% glycerol/TBS medium, after its appropriate preparation, and observe by excitation at 450-490 nm, using a dichromatic mirror at 510 nm and a long-pass filter at 515 nm (Evangelou et al., 2017).
- 4.8 Perform all incubations in dark chambers to avoid exsiccation of the material and exposure to light.
- 4.9 Incubation of cells with 70% EthOH just before addition of SenTraGor™ reagent is an essential step since SenTraGor™ is diluted in pure EthOH. Otherwise it will not be able to penetrate the cell and staining will not be successful.
- 4.10 This step is crucial to avoid evaporation of the dye.



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- 4.11 Absence of staining with SenTraGor™ reagent *per se* within 5-8 minutes does not always indicate that the sample is negative for senescence. From our experience we suggest to proceed with the DAB visualization reaction. In many cases, despite it was challenging to detect the positive granules after SenTraGor™ reagent staining, we clearly detected positive (dark brown) senescent cells after completion of the DAB reaction. The addition of the chromogenic assay increases dramatically the sensitivity of the method.
- 4.12 Intracellular light blue staining can occasionally be observed when the SenTraGor™ reagent is used, and should always be taken into consideration.
- 4.13 Omission of the SenTraGor™ reagent should always be performed as a negative control experiment.
- 4.14 This step is crucial to remove and estimate “background dirt” and clean cover glass and slides using soft paper.
- 4.15 The addition of the primary anti-biotin antibody is essential to obtain a consistent and specific signal. Omission of the primary anti-biotin antibody should always serve as negative control.
- 4.16 Incubation with solutions (BSA, blocking medium or corresponding sera) that block non-specific antibody staining is optional.
- 4.17 In case of using SenTraGor™ reagent *per se* in cells use 0.1% Nuclear Fast Red as counterstain.
- 4.18 Instead of a DAB detection method you can use AP (alkaline phosphatase) detection assay, without the execution of step 3.2.